

## Hematologic Disorders, Leukemia/Lymphoma; Flow Hold, Varies

**Test ID:** HLLFH

**Explanation:** Effective immediately, additional information will be added to the specimen requirements and testing algorithm to include spinal fluid as an acceptable source.

### Current Testing Algorithm

This test is designed to delay the start of leukemia/lymphoma immunophenotyping until the preliminary assessment is completed. Specimens are held in the laboratory until noon (12 p.m. Central time) 2 days after the collection date. For testing to be canceled, the client must call 800-533-1710. The testing process will be initiated and fully charged if no notification is received within this time period. To expedite the beginning of testing, call 800-533-1710.

### New Testing Algorithm

This test is designed to delay the start of leukemia/lymphoma immunophenotyping until the preliminary assessment is completed. **CSF specimens are held in the laboratory until noon (12 p.m. CST) 1 day after the collection date.** All other specimen types are held in the laboratory until noon (12 p.m. CST) 2 days after the collection date. For testing to be canceled, the client must call 800-533-1710. The testing process will be initiated and fully charged if no notification is received within this time period. To expedite the beginning of testing, call 800-533-1710.

### Current Specimen Required

**Submit only 1 of the following specimens:**

**Specimen Type:** Whole blood

**Container/Tube:**

**Preferred:** Yellow top (ACD solution A or B)

**Acceptable:** Lavender top (EDTA) or Green top (sodium heparin)

**Specimen Volume:** 10 mL

**Slides:** If possible, include 5- to 10-unstained blood smears, **must be labeled with two unique identifiers.**

**Collection Instructions:**

1. Send whole blood specimen in original tube. **Do not aliquot.**

2. Label specimen as blood.

**Specimen Stability Information:** Ambient < or =4 days/Refrigerated < or =4 days

**Specimen Type:** Bone marrow

**Container/Tube:**

### New Specimen Required

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**Specimen Type:** Whole blood

**Container/Tube:**

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**Acceptable:** Lavender top (EDTA) or Green top (sodium heparin)

**Specimen Volume:** 10 mL

**Slides:** If possible, include 5- to 10-unstained blood smears, **must be labeled with two unique identifiers.**

**Collection Instructions:**

1. Send whole blood specimen in original tube. **Do not aliquot.**

2. Label specimen as blood.

**Specimen Stability Information:** Ambient < or =4 days/Refrigerated < or =4 days

**Specimen Type:** Bone marrow

**Container/Tube:**

**Preferred:** Yellow top (ACD solution A or B)

**Acceptable:** Lavender top (EDTA) or green top (sodium heparin)

**Specimen Volume:** 1 to 5 mL

**Slides:** If possible, include 5- to 10-unstained bone marrow aspirate smears, which **must be labeled with two unique identifiers**

**Collection Instructions:**

1. Submission of bilateral specimens is not required.
2. Send bone marrow specimen in original tube. **Do not aliquot.**
3. Label specimen as bone marrow.

**Specimen Stability Information:** Ambient < or =4 days/Refrigerated < or =4 days

**Specimen Type:** Fluid

**Sources:** Serous effusions, pleural, pericardial, or abdominal (peritoneal fluid)

**Container/Tube:** Body fluid container

**Specimen Volume:** 20 mL

**Collection Instructions:**

1. If possible, fluids should be anticoagulated with heparin (1 U/mL of fluid).
2. Label specimen with fluid type.

**Specimen Stability Information:**

Refrigerated/Ambient < or =4 days

**Additional Information:** The volume of fluid necessary to phenotype the lymphocytes or blasts in serous effusions depends upon the cell count in the specimen. Usually, 20 mL of pleural or peritoneal fluid is sufficient. Smaller volumes can be used if there is a high cell count.

**Specimen Type:** Tissue

**Supplies:** Hank's Solution (T132)

**Container/Tube:** Sterile container with 15 mL of tissue culture medium (eg, Hank's balanced salt solution, RPMI, or equivalent)

**Specimen Volume:** 5 mm(3) or larger biopsy

**Collection Instructions:**

1. Send intact specimen (**do not mince**)
2. Specimen cannot be fixed.

**Specimen Stability Information:** Ambient < or =4 days/Refrigerated < or =4 days

**Preferred:** Yellow top (ACD solution A or B)

**Acceptable:** Lavender top (EDTA) or green top (sodium heparin)

**Specimen Volume:** 1 to 5 mL

**Slides:** If possible, include 5- to 10-unstained bone marrow aspirate smears, which **must be labeled with two unique identifiers**

**Collection Instructions:**

1. Submission of bilateral specimens is not required.
2. Send bone marrow specimen in original tube. **Do not aliquot.**
3. Label specimen as bone marrow.

**Specimen Stability Information:** Ambient < or =4 days/Refrigerated < or =4 days

**Note:** A fresh (less than 4 days post-collection), unfixed, nonembedded bone marrow core biopsy, bone or bone lesion is acceptable as an equivalent source for bone marrow aspirate for this test **only in the event of a dry tap** during the bone marrow harvesting procedure. Indicate "dry tap" in performing lab notes or paperwork when submitting this specimen type.

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**Collection Instructions:**

1. Send intact specimen (**do not mince**)
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**Specimen Stability Information:** Ambient < or =4 days/Refrigerated < or =4 days

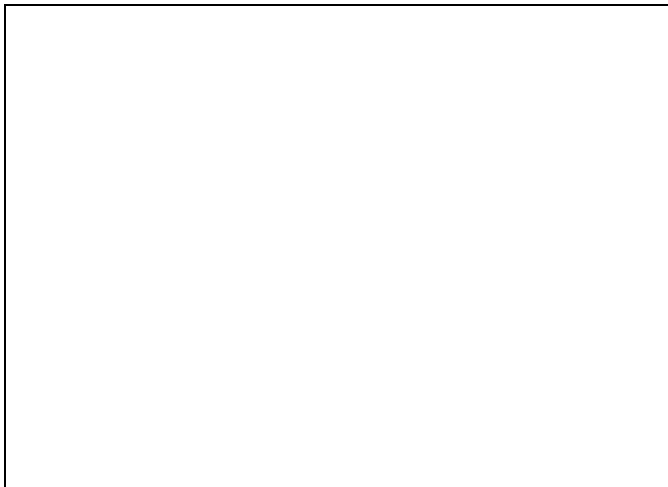
**Specimen Type:** Spinal fluid

**Container/Tube:** Sterile vial

**Specimen Volume:** 1 to 1.5 mL

**Collection Instructions:**

1. An original cytospin preparation (preferably unstained) should be included with the spinal fluid



specimen so correlative morphologic evaluation can occur.

2. Label specimen as spinal fluid.

**Specimen Stability Information:** Refrigerated 4 days/Ambient 4 days

**Additional Information:** The volume of spinal fluid necessary to phenotype the lymphocytes or blasts depends upon the cell count in the specimen. A cell count should be determined and submitted with the specimen. Usually, 1 to 1.5 mL of spinal fluid is sufficient. Smaller volumes can be used if there is a high cell count. If cell count is less than 10 cells/mcL, a larger volume of spinal fluid may be required. When cell counts drop below 5 cells/mcL, the immunophenotypic analysis may not be successful.

**Questions**

Contact Melissa Lonzo, Laboratory Resource Coordinator, at 800-533-1710.